

Amendments to the Specification:

1. Please replace the second last paragraph on page 16 with the following paragraph:

E1 --In another embodiment, the Family B DNA polymerase comprises a serine to asparagene asparagine mutation at a site corresponding to S651 of SEQ ID NO: 2.--

2. Please replace the first paragraph on page 7 with the following paragraph:

E2 -- Family B DNA polymerases exhibit substantially different structure compared to Family A DNA polymerases, with the exception of the position of acidic residues involved in catalysis in the so-called palm domain (Wang et al., 1997, Cell 89:1087; Hopfner et al., 1999, Proc. Natl. Acad. Sci. USA 96:3600). The unique structure of Family B DNA polymerases may permit a completely different spectrum of interactions with nucleotide analogs, perhaps allowing utilization of analogs which are unsuitable for use with Family A DNA polymerases due to structural constraints. Thermostable Family B DNA polymerases have been identified in hyperthermophilic archaea. These organisms grow at temperatures higher than ~~904~~ 91°C and their enzymes demonstrate greater thermostability (Mathur et al., 1992, Strategies 5:11) than the thermophilic eubacterial Family A DNA polymerases. Family B polymerases from hyperthermophilic archaea may be well suited starting substrates for modification(s) to reduce discrimination against non-conventional nucleotides.--

3. Please replace the last paragraph starting from line 11 on page 8 to line 10 on page 9 with the following paragraph:

E3 -- Mutagenesis studies done in Family B DNA polymerases also implicate the region containing the analogous Y in region II in dNTP incorporation and ribose selectivity. Mutations at the corresponding Y865 in human DNA polymerase α affect polymerase fidelity and sensitivity to dNTP nucleotide inhibitors such as AZT-TP, which has a bulky 3'-azido group in place of the 3'-OH group, BuPdGTP, which contains a butylphenyl group attached to the amino group at the C-2 position in the guanine base of dGTP (resulting in a bulkier and more hydrophobic purine base nucleotide) and aphidicolin, a competitive inhibitor of pyrimidine

E3
deoxynucleotide triphosphate. Interestingly, the mutants showed no difference in their uptake of ddCTP (Dong et al., 1993, J. Biol. Chem. 268: 26143 24163). Additionally, mutants of bacteriophage T4 DNA polymerase, which have converted L412 to methionine (M) or isoleucine (I) just one amino acid before the analogous Y (Y411), show extreme and mild sensitivity, respectively, to the inorganic pyrophosphate analog phosphonoacetic acid (PAA). Alterations in PAA sensitivity have been shown to predict polymerase interactions with nucleotide analogs. L412 in T4 DNA polymerase corresponds to L410 in *Thermococcus* species JDF-3 DNA polymerase. The L412M T4 DNA polymerase mutant was inhibited with 50-fold less ddGTP than wild-type polymerase while the K_m s for dGTP was similar. As stated by the authors in that study, "[d]espite the sensitivity of the L412M DNA polymerase to ddGTP, there was no difference found in the incorporation of ddNTPs by wild-type and L412M DNA polymerase." (Reha-Krantz et al., 1993, J. Virol. 67:60). In bacteriophage ϕ 29, mutations in region II (LYP where Y is analogous to *Thermococcus* species JDF3 DNA polymerase Y409) produce mixed results when challenged with PAA; P255S was hypersensitive to PAA while L253V was shown to be less sensitive than the wild-type enzyme (Blasco et al., 1993, J. Biol. Chem. 268: 24106). These data support the role of the LYP region (region II) in polymerase-nucleotide interactions, but improved incorporation of ddNTPs was not achieved in these references.--

4. Please replace the first full paragraph at page 56, from line 8 through line 20, with the following paragraph:

E4
-- DNA polymerases lacking 3'-5' exonuclease (proofreading) activity are preferred for applications requiring nucleotide analog incorporation (e.g., DNA sequencing) to prevent removal of nucleotide analogs after incorporation. The 3'-5' exonuclease activity associated with proofreading DNA polymerases can be reduced or abolished by mutagenesis. Sequence comparisons have identified three conserved motifs (exo I (DXE), II (NX₂₋₃(F/Y)D), III (YX₃D)) in the 3'-5' exonuclease domain of DNA polymerases (reviewed V. Derbyshire, J.K. Pinsonneault, and C.M. Joyce, Methods Enzymol. 262, 363 (1995)). Replacement of any of the conserved aspartic or glutamic acid residues with alanine has been

E4 shown to abolish the exonuclease activity of numerous DNA polymerases, including archaeal DNA polymerases such as Vent (H. Kong, R.B. Kucera, and W.E. Jack, J. Biol. Chem. 268, 1965 (1993)) and *Pfu* (Stratagene, unpublished). Conservative substitutions lead to reduced exonuclease activity, as shown for mutants of the archaeal 9° N-7 DNA polymerase (M.W. Southworth, H. Kong, R.B. Kucera, J. Ware, H. Jannasch, and F.B. Perler, Proc. Natl. Acad. Sci. 93, 5281 (1996)). --
